Direct Activation of Bmi1 by Twist1: Implications in Cancer Stemness, Epithelial-Mesenchymal Transition, and Clinical Significance

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Cancer stemness is a concept used to describe a minor population of cells (cancer stem cells-CSCs) residing in a tumor, which possess self-renewal properties and are resistant to chemo/radiation therapy. Epithelial-mesenchymal transition (EMT), a major mechanism of cancer metastasis, is a process which generates cells with stem-like properties. The relationship between cancer stemness and EMT is well documented but without detailed mechanistic explanation. Bmi1 belongs to the polycomb repressive complex 1 (PRC1) which maintains self-renewal and stemness. Recent results showed that Twist1, an EMT regulator, directly activates Bmi1 and these two molecules function together to mediate cancer stemness and EMT. These results provide a molecular explanation of the relationship between cancer stemness and EMT. Bmi1 is frequently overexpressed in various types of human cancers and can con-

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fer drug resistance. Twist1 is also overexpressed in various human cancers with prognostic significance. The functional interdependence between Twist1 and Bmi1 provides a fresh insight into the molecular mechanism of EMT-induced cancer stemness. Further investigation of the mechanisms mediating EMT and cancer stemness will be helpful in the management and treatment of metastatic cancers. *(Chang Gung Med J 2011;34:229-38)*

Key words: Epithelial-mesenchymal transition, cancer stemness, Twist1, Bmi1

Cancer stemness is a concept recently proposed to

explain cancer cells' resistance to conventional
 C^2 chemo/radiation therapy.(1) Cancer stem cells (CSCs) are described as a small percentage of cells residing in a tumor, which are able to self-renew and have stem-like properties.⁽¹⁻³⁾ Stem-like properties are monitored by different assays such as staining of surface markers, *in vitro* sphere formation, resistance to chemotherapeutic agents or radiation, *in vivo* tumorinitiating capability, and other assays. $(1,4)$

Epithelial-mesenchymal transition (EMT) is an important process by which epithelial cells are converted to mesenchymal cells during embryonic development.(5-7) This process involves loss of cell polarity, decrease in cell-to-cell adhesion, and gain of migration ability.(5-7) EMT is also the critical event

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for tumor metastasis and organ fibrosis.⁽⁵⁻⁷⁾ Repression of epithelial markers (e.g., E-cadherin, plakoglobin and desmoplakin) and upregulation of mesenchymal markers (e.g., vimentin, fibronectin and N-cadherin) are the typical marker changes observed during the EMT process.(5-7) Different transcription factors such as Snail (also known as SNAI1), Slug (also known as SNAI2), Zeb1 (also known as TCF8 and δEF1), SIP1 (also known as Zeb2 and ZFXH1B), E47 (also known as TCF3), and Twist1 are termed "EMT regulators" since they were shown to regulate $EMT₀⁽⁸⁻¹³⁾$ In spite of the demonstration of the role of EMT in embryonic development, cancer metastasis, and organ fibrosis, it is unknown whether EMT plays a significant role in other aspects of cell biology.

This review summarizes recent findings in the relationship between cancer stemness and epithelialmesenchymal transition, the regulation of *Bmi1* by Twist1 and its significance in cancer stemness, and the role of Bmi1 and Twist1 in contributing to various types of human cancers.

The relationship between cancer stem cells and epithelial-mesenchymal transition

The cancer stem cells possess the ability to selfrenew and generate secondary tumors, which is described as a "tumor-initiating ability" and is best assayed by *in vivo* limiting dilution assays.⁽⁴⁾ It is hypothesized that solid tumors are hierarchically organized and sustained by cancer stem cells.⁽¹⁴⁾ For example, after treatment of breast cancer, the surviving residual tumor cells may be enriched for subpopulations of cells (e.g. CD44+/CD24- or low) with both tumor-initiating and mesenchymal features.⁽¹⁵⁾ Different developmental pathways such as hedgehog, epidermal growth factor receptor (EGFR), Wnt/βcatenin, Notch, polycomb (Bmi1), stromal cell derived factor-1 (SDF-1)/chemokine receptor-4 (CXCR4), PTEN, BMP, and TGF-β were shown to be associated with tumor-initiating abilities.⁽¹⁶⁾ In addition, various cancer subtypes may have different subsets of tumor-initiating cells. For example, CSCs could be isolated or monitored by different cell-surface marker profiles in various types of human cancers (e.g. CD133 in brain, colon, pancreas, lung, and ovarian cancers; CD44 in breast and head and neck cancers).(4) CSCs are associated with a specific state of differentiation (e.g. mesenchymal features). (17)

The EMT process in tumor cells usually results in cells becoming more invasive, metastatic, and drug resistant, which will lead to the subsequent demise of cancer patients.^{$(5-7)$} It is well documented that EMT will induce tumor progression and aggressiveness.⁽⁵⁻⁷⁾ EMT-derived cells exhibit multi-lineage differentiation potential similar to mesenchymal stem cells.(18) However, the mechanisms delineating the connection between EMT and cancer stemness are largely unknown. Recent evidences suggest that the process of EMT generates cells with stem-like properties.(19) The earliest example is the generation of proliferative human islet precursor cells during EMT.⁽²⁰⁾ Loss of p21CIP1 is associated with the generation of breast cancer stem cell properties.(21) Other examples include the demonstration that the EMT process generates stem-like properties in breast cancer cells.(22,23) Since cancer stem cells may have characteristics different from the original tumor cells, or the tumor cells sensitive to chemo/radiation therapy, the link between EMT and cancer stemness provides the explanation that EMT induces tumor progression through induction of cancer stemness.(24-26)

Bmi1, a polycomb protein, regulates and maintains stemness features

Polycomb group (PcG) proteins are chromatin modifiers involved in the maintenance of embryonic and adult stem cells and cancer formation.⁽²⁷⁻³³⁾ Polycomb group proteins are multimeric transcriptional repressor complexes including polycombrepressive complex 1 (PRC1) and polycomb-repressive complex 2 (PRC2).⁽²⁷⁻³³⁾ Polycomb group proteins can occupy the promoters of developmental regulators, and silencing of these genes confers stemness in a PcG-dependent manner.(27-33)

Bmi1 is a member of polycomb-repressive complex 1 (PRC1) which maintains chromatin silencing.(34) Bmi1 was first shown to collaborate with c-Myc to promote lymphomagenesis and regulate cell proliferation and senescence through inhibiting the *INK4A* locus, demonstrating its role as an oncogene.(35,36) Bmi1 was subsequently shown to be required to maintain normal and leukemic hematopoietic stem cells and was essential in the lineage specification and multipotency of hematopoietic stem and progenitor cells.^{$(37-39)$} In addition, Bmi1 was shown to be involved in the self-renewal of mammary epithelium, neuronal, pancreatic (includ-

ing β-cell), and intestinal cells through repressing the *INK4A-ARF* locus.⁽⁴⁰⁻⁴⁷⁾ However, repression of *INK4A-ARF* by Bmi1 is dependent on the polycombrepressive complex 2 (PRC2).⁽⁴⁸⁾ After PRC2 binds to the promoters of target genes, EZH2 (a member of PRC2 with histone H3 methyltransferase activity) methylates lysine 27 of histone H3 (H3K27).^(49,50) The trimethylated H3K27 (H3K27me3) is then recognized and bound by PRC1.⁽⁵¹⁾ Both PRC1 and PRC2 bind to the promoters of target genes to maintain their repression. Among the target genes of PRC1 and PRC2, repression of the *INK4A/ARF* locus is essential for PRC complexes to maintain stemness.(52,53)

Direct regulation of *Bmi1* **by Twist1: implications in cancer stemness induced by epithelialmesenchymal transition**

Twist1, a bHLH transcriptional factor, was first demonstrated for its critical role in the *Drosophila* mesoderm development.^(54,55) Twist1 governs cell movement and tissue reorganization during early embryogenesis and is a master regulator of gastrulation, mesoderm differentiation, and somatic muscles patterning and specification.⁽⁵⁶⁾ The critical role of Twist1 in cancer metastasis was recently demonstrated by the results of increased expression of Twist1 in human cancers, induction of EMT by Twist1, and the association of Twist1 with a more aggressive phenotype and a worse outcome. $(10,57)$ Twist1 expression is triggered by different upstream signaling pathways.⁽⁵⁸⁾ We previously demonstrated that hypoxia inducible factor-1 (HIF-1) directly regulates *Twist1* expression.⁽⁵⁹⁾ Recent results also identified a subpopulation of highly tumorigenic cells in head and neck squamous cell carcinoma (HNSCC) with stem-like properties and overexpressing Bmi1.(60) Due to the link between EMT and cancer stemness, we hypothesized that Twist1 may induce the expressions of stemness genes, resulting in the promotion of EMT and tumor-initiating ability. Through the screening of possible activation of different stemness genes, a tight correlation between Twist1 and Bmi1 was observed.(61) Different assays such as transient transfection, electrophoretic mobility shift assay (EMSA), and chromatin immunoprecipitation (ChIP) assays were subsequently performed to demonstrate the direct activation of *Bmi1* expression by Twist1. Overexpression of Twist1 or Bmi1 conferred stemlike properties and induced EMT in head and neck cancer cell lines. Bmi1 was critical for Twist1 induced stem-like properties and EMT since knockdown of Bmi1 in Twist1-overexpressing cells abolished both stem-like properties and EMT. Overexpression of Bmi1 alone could induce EMT. Twist1 was also critical for Bmi1-induced stem-like properties and EMT since knockdown of Twist1 in Bmi1-overexpressing cells reversed EMT and abolished stem-like properties. Quantitative chromatin immunoprecipitation (qChIP) assays were performed to test the binding of these two proteins on both *Ecadherin* and *p16INK4A* promoters when either Bmi1 or Twist1 was knocked down. The results showed the functional interdependence of Twist1 and Bmi1 to mediate stem-like properties and EMT since knockdown of either molecule caused the decreased binding of the other molecule on both promoters. Since repression of *E-cadherin* by Twist1 was not shown previously, (7) we further mapped three E-box sites located in the *E-cadherin* promoter responsible for Twist1-induced repression by mutagenesis analysis of these three E-box sites. Electrophoretic mobility shift assay (EMSA) followed by supershifting with the anti-Twist1 or anti-Bmi1-specific antibody showed the co-occupancy of the *E-cadherin* promoter by Twist1 and Bmi1. The essential role of EZH2 was also demonstrated using the assays mentioned above,(61) which was consistent with the reported result.(62) Chromatin immunoprecipitation assays showed the direct binding of Twist1 to the *Bmi1* promoter. Co-immunoprecipitation assay showed the interaction between Twist1 and Bmi1. Our results present the first molecular demonstration of simultaneous repression of both *E-cadherin* and *p16INK4A* expression by Twist1 (an EMT regulator) and Bmi1/EZH2 (components of the polycomb group proteins). These results provide one of the first molecular delineations of the link between cancer stemness and EMT.(61) Transcriptome profiling analysis also showed that head and neck cancer cell lines overexpressing Twist1 or Bmi1 had the transcriptome drifting to the mesenchymal stem cell signatures, but not drifting to the epithelial transcriptome signatures.⁽⁶¹⁾ This result is consistent with the reported result that cancer stem cells display mesenchymal features.(17) Finally, the important role of Bmi1 in cancer stemness is supported by the recent result that Bmi1 was critical in the maintenance of prostate cancer stem cells.⁽⁶³⁾

Cancer stemness, Bmi1, and drug resistance

Tumor- and metastasis-initiating cells usually develop treatment resistance, which is shown in recurrent ovarian cancer.⁽⁶⁴⁾ Activated CD8 T cells can stimulate mammary tumor cells to go through EMT and increase their tumor-initiating ability and chemotherapeutic resistance.(65) Bmi1 is shown to confer different kinds of resistance (radiation, 5-fluorouracil, docetaxel) in different cancers.⁽⁶⁶⁻⁶⁸⁾ Recruitment of the DNA damage response machinery is shown to cause Bmi1-induced radiation resistance.(69) The detailed molecular mechanisms of treatment resistance are still largely unknown. Future experiments to dissect the signaling pathways regulating different kinds of treatment resistance will require intensive investigation. Finally, the observation and concept of cancer stem cells could mimic the "minimal residual disease" constantly mentioned during the treatment course of certain leukemias.⁽⁷⁰⁾

Expression of Twist1 and Bmi1 and their contribution to clinical significance

Bmi1 overexpression is shown in various cancers such as chronic myeloid leukemia, multiple myeloma, head and neck squamous cell carcinoma, squamous cell carcinoma of the tongue, breast cancers, non-small cell lung cancer, hepatocellular carcinoma, gastric carcinoma, Ewing sarcoma, colon cancer, bladder cancer, esophageal cancer, cholangiocarcinoma, ovarian cancer, endometrial cancer, cervical cancer, and medulloblastoma.(71-95) Bmi1 cooperates with H-Ras to induce aggressive breast cancer. (96) Bmi1 collaborates with BCR-ABL or interacts with PLZF/RARA to mediate leukemic transformation.(97,98) Finally, Bmi1 induces apoptotic resistance through activation of the IKK-NF-kB pathway.⁽⁹⁹⁾ The prognostic impact of Twist1 was demonstrated in various cancers,(10,59,100-116) but the interdependence between Twist1 and Bmi1 has never been explored. The cooperative role between Twist1 and Bmi1 was demonstrated in HNSCC cases since only co-overexpression of both proteins correlates with repression of *E-cadherin* and *p16INK4A* and the worst prognosis of HNSCC patients.⁽⁶¹⁾ Patients expressing either Twist1 or Bmi1 alone have a better prognosis than those co-expressing both proteins. This observation further strengthens the model that Twist1 and Bmi1

interdependently promote EMT and cancer stemness, resulting in an aggressive tumor behavior and a dismal outcome in HNSCC.⁽⁶¹⁾ Further confirmation of this functional interdependence will require examination of more tumor samples from various tumor types.

Conclusions and future perspectives

Different mechanisms such as chromatin modification (e.g. promoter hypermethylation) and recruitment of chromatin modifiers such as HDAC1/HDAC2, AJUBA/PRMT5, or PRC2 by EMT regulators were shown to mediate *E-cadherin* repression.(117-119) However, there was no previous demonstration of the involvement of PRC1 complex in the repression of *E-cadherin*. In spite of the repeated demonstrations that *p16INKA* is regulated by Bmi1,(35,36,47) the involvement of an EMT regulator in the repression of *p16INK4A* was not shown. Our results are the first demonstration of the requirement of an EMT regulator and PRC1/2 complexes to simultaneously repress *E-cadherin* and *p16INK4A*. The simultaneous requirement of transcription regulators and chromatin modification complexes (PRC1 and PRC2 in our case) to mediate E-cadherin and *p16INK4A* repression provides a mechanistic example of the relationship between EMT and cancer stemness.

Bmi1 is well documented to maintain stemness.(34,37-39) Twist1 and Twist2 also override oncogene-induced premature senescence in cancer cells by inhibiting the activity of *p16INK4A* and *p21CIP1*. (120) From our results, it appears that Bmi1 acts together with Twist1 to carry out multiple functions, including EMT induction and escape from oncogene-induced premature senescence. Other functions such as induction of telomerase activity, inhibition of TGF-β signaling, and repression of PTEN tumor suppressor were also mediated by Bmi1,⁽¹²¹⁻¹²³⁾ which may contribute to Bmi1-mediated functions.

In conclusion, the relationship between cancer stemness and EMT is well established. Our demonstration that Twist1 activates Bmi1 and both molecules function interdependently to mediate cancer stemness and EMT provide a molecular delineation of the relationship between cancer stemness and EMT. Investigation of the regulation of Bmi1 or other stemness genes through different mechanisms should be the subject which needs immediate attention to further explore this relationship. The information obtained from these investigations will be valuable for the management and treatment of metastatic cancers.

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REFERENCES

- 1. Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. Nature 2001;414:105- 11.
- 2. Gupta PB, Chaffer CL, Weinberg RA. Cancer stem cells: mirage or reality? Nat Med 2009;15:1010-2.
- 3. Marotta LL, Polyak K. Cancer stem cells: a model in the making. Curr Opin Genet Dev 2009;19:44-50.
- 4. O'Brien CA, Kreso A, Jamieson CH. Cancer stem cells and self-renewal. Clin Cancer Res 2010;16:3113-20.
- 5. Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. J Clin Invest 2009;119:1420-8.
- 6. Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelialmesenchymal transitions in development and disease. Cell 2009;139:871-90.
- 7. Yang J, Weinberg RA. Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. Dev Cell 2008;14:818-29.
- 8. Cano A, Pérez-Moreno MA, Rodrigo I, Locascio A, Blanco MJ, del Barrio MG, Portillo F, Nieto MA. The transcription factor snail controls epithelial-mesenchymal transitions by repressing E-cadherin expression. Nat Cell Biol 2000;2:76-83.
- 9. Hajra KM, Chen DY, Fearon ER. The SLUG zinc-finger protein represses E-cadherin in breast cancer. Cancer Res 2002;62:1613-8.
- 10. Yang J, Mani S, Donaher J, Ramaswamy S, Itzykson R, Come C, Savagner P, Gitelman I, Richardson A, Weinberg R. Twist, a master regulator of morphogenesis, plays an essential role in tumor metastasis. Cell 2004;117:927-39.
- 11. Grooteclaes ML, Frisch SM. Evidence for a function of CtBP in epithelial gene regulation and anoikis. Oncogene 2000;19:3823-8.
- 12. Comijn J, Berx G, Vermassen P, Verschueren K, van

Grunsven L, Bruyneel E, Mareel M, Huylebroeck D, van Roy F. The two-handed E box binding zinc finger protein SIP1 downregulates E-cadherin and induces invasion. Mol Cell 2001;7:1267-78.

- 13. Pérez-Moreno MA, Locascio A, Rodrigo I, Dhondt G, Portillo F, Nieto MA, Cano A. A new role for E12/E47 in the repression of E-cadherin expression and epithelialmesenchymal transitions. J Biol Chem 2001;276:27424- 31.
- 14. Polyak K, Weinberg RA. Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. Nat Rev Cancer 2009;9:265-73.
- 15. Blick T, Hugo H, Widodo E, Waltham M, Pinto C, Mani SA, Weinberg RA, Neve RM, Lenburg ME, Thompson EW. Epithelial mesenchymal transition traits in human breast cancer cell lines parallel the CD44(hi/)CD24 (lo/-) stem cell phenotype in human breast cancer. J Mammary Gland Biol Neoplasia 2010;15:235-52.
- 16. Mimeault M, Batra SK. New advances on critical implications of tumor- and metastasis-initiating cells in cancer progression, treatment resistance and disease recurrence. Histol Histopathol 2010;25:1057-73.
- 17. Creighton CJ, Li X, Landis M, Dixon JM, Neumeister VM, Sjolund A, Rimm DL, Wong H, Rodriguez A, Herschkowitz JI, Fan C, Zhang X, He X, Pavlick A, Gutierrez MC, Renshaw L, Larionov AA, Faratian D, Hilsenbeck SG, Perou CM, Lewis MT, Rosen JM, Chang JC. Residual breast cancers after conventional therapy display mesenchymal as well as tumor-initiating features. Proc Natl Acad Sci USA 2009;106:13820-5.
- 18. Battula VL, Evans KW, Hollier BG, Shi Y, Marini FC, Ayyanan A, Wang RY, Brisken C, Guerra R, Andreeff M, Mani SA. Epithelial-mesenchymal transition-derived cells exhibit multilineage differentiation potential similar to mesenchymal stem cells. Stem Cells 2010;28:1435-45.
- 19. Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, Brooks M, Reinhard F, Zhang CC, Shipitsin M. Campbell LL, Polyak K, Brisken C, Yang J, Weinberg RA. The epithelial-mesenchymal transition generates cells with properties of stem cells. Cell 2008;133:704-15.
- 20. Gershengorn MC, Hardikar AA, Wei C, Geras-Raaka E, Marcus-Samuels B, Raaka BM. Epithelial-to-mesenchymal transition generates proliferative human islet precursor cells. Science 2004;306:2261-4.
- 21. Liu M, Casimiro MC, Wang C, Shirley LA, Jiao X, Katiyar S, Ju X, Li Z, Yu Z, Zhou J, Johnson M, Fortina P, Hyslop T, Windle JJ, Pestell RG. p21CIP1 attenuates Rasand c-Myc-dependent breast tumor epithelial mesenchymal transition and cancer stem cell-like gene expression in vivo. Proc Natl Acad Sci USA 2009;106:19035-9.
- 22. Morel AP, Lièvre M, Thomas C, Hinkal G, Ansieau S, Puisieux A. Generation of breast cancer stem cells through epithelial-mesenchymal transition. PLoS One 2008;3:e2888.
- 23. Creighton CJ, Chang JC, Rosen JM. Epithelial-mesenchy-

mal transition (EMT) in tumor-initiating cells and its clinical implications in breast cancer. J Mammary Gland Biol Neoplasia 2010;15:253-60.

- 24. Hollier BG, Evans K, Mani SA. The epithelial-to-mesenchymal transition and cancer stem cells: a coalition against cancer therapies. J Mammary Gland Biol Neoplasia 2009;14:29-43.
- 25. Kong D, Banerjee S, Ahmad A, Li Y, Wang Z, Sethi S, Sarkar FH. Epithelial to mesenchymal transition is mechanistically linked with stem cell signatures in prostate cancer cells. PLoS One 2010;5:e12455.
- 26. Ouyang G, Wang Z, Fang X, Liu J, Yang CJ. Molecular signaling of the epithelial to mesenchymal transition in generating and maintaining cancer stem cells. Cell Mol Life Sci 2010;67:2605-18.
- 27. Valk-Lingbeek ME, Bruggeman SW, van Lohuizen M. Stem cells and cancer: the Polycomb connection. Cell 2004;118:409-18.
- 28. Gil J, Bernard D, Peters G. Role of polycomb group proteins in stem cell self-renewal and cancer. DNA Cell Biol 2005;24:117-25.
- 29. Buszczak M, Spradling AC. Searching chromatin for stem cell identity. Cell 2006;125:233-6.
- 30. Sparmann A, van Lohuizen M. Polycomb silencers control cell fate, development and cancer. Nat Rev Cancer 2006;6:846-56.
- 31. Rajasekhar VK, Begemann M. Concise review: roles of polycomb group proteins in development and disease: a stem cell perspective. Stem Cells 2007;25:2498-510.
- 32. Pietersen AM, van Lohuizen M. Stem cell regulation by polycomb repressors: postponing commitment. Curr Opin Cell Biol 2008;20:201-7.
- 33. Konuma T, Oguro H, Iwama A. Role of the polycomb group proteins in hematopoietic stem cells. Dev Growth Diff 2010;52:505-15.
- 34. Park IK, Morrison SJ, Clarke MF. Bmi1, stem cells, and senescence regulation. J Clin Invest 2004;113:175-9.
- 35. Jacobs JJ, Scheijen B, Voncken JW, Kieboom K, Berns A, van Lohuizen M. Bmi-1 collaborates with c-Myc in tumorigenesis by inhibiting c-Myc-induced apoptosis via INK4a/ARF. Genes Dev 1999;13:2678-90.
- 36. Jacobs JJ, Kieboom K, Marino S, Depinho R, van Lohuizen M. The oncogene and Polycomb-group gene Bmi1 regulates cell proliferation and senescence through the ink4a locus. Nature 1999;397:164-8.
- 37. Lessar J, Sauvageau G. Bmi-1 determines the proliferative capacity of normal and leukaemic stem cells. Nature 2003;423:255-60.
- 38. Park IK, Qian D, Kiel M, Becker MW, Pihalja M, Weissman IL, Morrison SJ, Clarke MF. Bmi-1 is required for maintenance of adult self-renewing haematopoietic stem cells. Nature 2003;423:302-5.
- 39. Iwama A, Oguro H, Negishi M, Kato Y, Morita Y, Tsukui H, Ema H, Kamijo T, Katoh-Fukui Y, Koseki H, van Lohuizen M, Nakauchi H. Enhanced self-renewal of

hematopoietic stem cells mediated by the polycomb gene product Bmi-1. Immunity 2004;21:843-51.

- 40. Molofsky AV, Pardal R, Iwashita T, Park IK, Clarke MF, Morrison SJ. Bmi-1 dependence distinguishes neural stem cell self-renewal from progenitor proliferation. Nature 2003;425:962-7.
- 41. Bruggeman SW, Valk-Lingbeek ME, van der Stoop, PP, Jacobs JJ, Kieboom K, Tanger E, Hulsman D, Leung C, Arsenijevic Y, Marino S, van Lohuizen M. Ink4a and Arf differentially affect cell proliferation and neural stem cell self-renewal in Bmi1-deficient mice. Genes Dev 2005;19:1438-43.
- 42. Fasano CA, Dimos JT, Ivanova NB, Lowry N, Lemischka IR, Temple S. shRNA knockdown of Bmi-1 reveals a critical role for p21-Rb pathway in NSC self-renewal during development. Cell Stem Cell 2007;1:87-99.
- 43. Fasano CA, Phoenix TN, Kokovay E, Lowry N, Elkabetz Y, Dimos JT, Lemischka IR, Studer L, Temple S. Bmi-1 cooperates with Foxg1 to maintain neural stem cell selfrenewal in the forebrain. Genes Dev 2009;23:561-74.
- 44. Sangiorgi E, Capecchi MR. Bmi1 is expressed in vivo in intestinal stem cells. Nat Genet 2008;40:915-20.
- 45. Pietersen AM, Evers B, Prasad AA, Tanger E, Cornelissen-Steijger P, Jonkers J, van Lohuizen M. Bmi1 regulates stem cells and proliferation and differentiation of committed cells in mammary epithelium. Curr Biol 2008;18:1094-9.
- 46. Sangiorgi E, Capecchi MR. Bmi1 lineage tracing identifies a self-renewing pancreatic acinar cell subpopulation capable of maintaining pancreatic organ homeostasis Proc Natl Acad Sci USA 2009;106:7101-6.
- 47. Dhawan S, Tschen SI, Bhushan A. Bmi-1 regulates the Ink4a/Arf locus to control pancreatic beta-cell proliferation. Genes Dev 2009;23:906-11.
- 48. Pereira CF, Piccolo FM, Tsubouchi T, Sauer S, Ryan NK, Bruno L, Landeira D, Santos J, Banito A, Gil J, Hoseki H, Merkenschlager M, Fisher AG. ESCs require PRC2 to direct the successful reprogramming of differentiated cells toward pluripotency. Cell Stem Cell 2010;6:547-56.
- 49. Czermin B, Melfi R, McCabe D, Seitz V, Imhof A, Pirrotta V. Drosophila Enhancer of Zeste/ESC complexes have a histone H3 methyltransferase activity that marks chromosomal Polycomb sites. Cell 2002;111:185-96.
- 50. Müller J, Hart CM, Francis NJ, Vargas ML, Sengupta A, Wild B, Miller EL, O'Connor MB, Kingston RE, Simon JA. Histone methyltransferase activity of a *Drosophila* Polycomb group repressor complex. Cell 2002;111:197- 208.
- 51. Min J, Zhang Y, Xu RM. Structural basis for specific binding of Polycomb chromodomain to histone H3 methylated at Lys 27. Genes Dev 2003;17:1823-8.
- 52. Gil J, Peters G. Regulation of the INK4b-ARF-INK4a tumour suppressor locus: all for one or one for all. Nat Rev Mol Cell Biol 2006;7:667-77.
- 53. Bracken AP, Kleine-Kohlbrecher D, Dietrich N, Pasini D,

Gargiulo G, Beekman C, Theilgaard-Mönch K, Minucci S, Porse BT, Marine JC, Hansen KH, Helin K. The Polycomb group proteins bind throughout the INK4A–ARF locus and are disassociated in senescent cells. Genes Dev 2007;21:525-30.

- 54. Nusslein-Volhard C, Wieschaus E, Kluding, H. Mutations affecting the pattern of the larval cuticle in Drosophila melanogaster. 1. Zygotic loci on the 2nd chromosome. Roux's Arch Dev Biol 1984;193:267-82.
- 55. Castanon I, Baylies MK. Twist in fate: evolutionary comparison of Twist structure and function. Gene 2002;287: 11-22.
- 56. Furlong EE, Andersen EC, Null B, White KP, Scott MP. Patterns of gene expression during Drosophila mesoderm development. Science 2001;293:1629-33.
- 57. Yang J, Mani SA, Weinberg RA. Exploring a new twist on tumor metastasis. Cancer Res 2006;66:4549-52.
- 58. Yang MH, Wu KJ. TWIST activation by hypoxia inducible factor-1 (HIF-1): implications in metastasis and development. Cell Cycle 2008;7:2090-6.
- 59. Yang MH, Wu MZ., Chiou SH, Chen PM, Chang SY, Liu CJ, Teng SC, Wu KJ. Direct regulation of TWIST by HIF-1alpha promotes metastasis. Nat Cell Biol 2008;10:295-305.
- 60. Prince ME, Sivanandan R, Kaczorowski A, Wolf GT, Kaplan MJ, Dalerba P, Weissman IL, Clarke MF, Ailles LE. Identification of a subpopulation of cells with cancer stem cell properties in head and neck squamous cell carcinoma. Proc Natl Acad Sci USA 2007;104:973-8.
- 61. Yang MH, Hsu DS, Wang HW, Yang WH, Kao SY, Tzeng CH, Tai SK, Chang SY, Lee OK, Wu KJ. Bmi1 is essential in Twist1-induced epithelial-mesenchymal transition. Nat Cell Biol 2010;12:982-92.
- 62. Cao Q, Yu J, Dhanasekaran SM, Kim JH, Mani RS, Tomlins SA, Mehra R, Laxman B, Cao X, Yu J, Kleer CG, Narambally S, Chinnaiyan AM. Repression of E-cadherin by the polycomb group protein EZH2 in cancer. Oncogene 2008;27:7274-84.
- 63. Lukacs RU, Memarzadeh S, Wu H, Witte ON. Bmi-1 is a crucial regulator of prostate stem cell self-renewal and malignant transformation. Cell Stem Cell 2010;7:682-93.
- 64. Ahmed N, Abubaker K, Findlay J, Quinn M. Epithelial mesenchymal transition and cancer stem cell-like phenotypes facilitate chemoresistance in recurrent ovarian cancer. Curr Cancer Drug Targets 2010;10:268-78.
- 65. Reiman JM, Knutson KL, Radisky DC. Immune promotion of epithelial-mesenchymal transition and generation of breast cancer stem cells. Cancer Res 2010;70:3005-8.
- 66. Qin L, Zhang X, Zhang L, Feng Y, Weng GX, Li MZ, Kong QL, Qian CN, Zeng YX, Zeng MS, Liao DF, Song LB. Downregulation of BMI-1 enhances 5-fluorouracilinduced apoptosis in nasopharyngeal carcinoma cells. Biochem Biophys Res Commun 2008;371:531-5.
- 67. Alajez NM, Shi W, Hui AB, Yue S, Ng R, Lo KW, Bastianutto C, O'Sullivan B, Gullane P, Liu FF. Targeted

depletion of BMI1 sensitizes tumor cells to P53-mediated apoptosis in response to radiation therapy. Cell Death Differ 2009;16:1469-79.

- 68. Crea F, Duhagon Serrat MA, Hurt EM, Thomas SB, Danesi R, Farrar WL. BMI1 Silencing enhances docetaxel activity and impairs antioxidant response in prostate cancer. Int J Cancer 2010;128:1946-54.
- 69. Facchino S, Abdouh M, Chatoo W, Bernier G. BMI1 confers radioresistance to normal and cancerous neural stem cells through recruitment of the DNA damage response machinery. J Neurosci 2010;30:10096-111.
- 70. Raimondi C, Gianni W, Cortesi E, Gazzaniga P. Cancer stem cells and epithelial-mesenchymal transition: revisiting minimal residual disease. Curr Cancer Drug Targets 2010;10:496-508.
- 71. Bhattacharyya J, Mihara K, Yasunaga S, Tanaka H, Hoshi M, Takihara Y, Kimura A. BMI-1 expression is enhanced through transcriptional and posttranscriptional regulation during the progression of chronic myeloid leukemia. Ann Hematol 2009;88:333-40.
- 72. Jagani Z, Wiederschain D, Loo A, He D, Mosher R, Fordjour P, Monahan J, Morrissey M, Yao YM, Lengauer C, Warmuth M, Sellers WR, Dorsch M. The Polycomb group protein Bmi-1 is essential for the growth of multiple myeloma cells. Cancer Res 2010;70:5528-38.
- 73. Häyry V, Mäkinen LK, Atula T, Sariola H, Mäkitie A, Leivo I, Keski-Säntti H, Lundin J, Haglund C, Hagström J. Bmi-1 expression predicts prognosis in squamous cell carcinoma of the tongue. Br J Cancer 2010;102:892-7.
- 74. Vormittag L, Thurnher D, Geleff S, Pammer J, Heiduschka G, Brunner M, Grasl MCh, Erovic BM. Coexpression of Bmi-1 and podoplanin predicts overall survival in patients with squamous cell carcinoma of the head and neck treated with radio (chemo) therapy. Int J Radiat Oncol Biol Phys 2009;73:913-8.
- 75. Kang MK, Kim RH, Kim SJ, Yip FK, Shin KH, Dimri GP, Christensen R, Han T, Park NH. Elevated Bmi-1 expression is associated with dysplastic cell transformation during oral carcinogenesis and is required for cancer cell replication and survival. Br J Cancer 2007;96:126-33.
- 76. Saeki M, Kobayashi D, Tsuji N, Kuribayashi K, Watanabe N. Diagnostic importance of overexpression of Bmi-1 mRNA in early breast cancers. Int J Oncol 2009;35:511-5.
- 77. Pietersen AM, Horlings HM, Hauptmann M, Langerød A, Ajouaou A, Cornelissen-Steijger P, Wessels LF, Jonkers J, van de Vijver MJ, van Lohuizen M. EZH2 and BMI1 inversely correlate with prognosis and TP53 mutation in breast cancer. Breast Cancer Res 2008;10:R109.
- 78. Becker M, Korn C, Sienerth AR, Voswinckel R, Luetkenhaus K, Ceteci F, Rapp UR. Polycomb group protein Bmi1 is required for growth of RAF driven nonsmall-cell lung cancer. PLoS One 2009;4:e4230.
- 79. Vrzalikova K, Skarda J, Ehrmann J, Murray PG, Fridman E, Kopolovic J, Knizetova P, Hajduch M, Klein J, Kolek V, Radova L, Kolar Z. Prognostic value of Bmi-1 onco-

protein expression in NSCLC patients: a tissue microarray study. J Cancer Res Clin Oncol 2008;134:1037-42.

- 80. Dovey JS, Zacharek SJ, Kim CF, Lees JA. Bmi1 is critical for lung tumorigenesis and bronchioalveolar stem cell expansion. Proc Natl Acad Sci USA 2008;105:11857-62.
- 81. Chiba T, Seki A, Aoki R, Ichikawa H, Negishi M, Miyagi S, Oguro H, Saraya A, Kamiya A, Nakauchi H, Yokosuka O, Iwama A. Bmi1 promotes hepatic stem cell expansion and tumorigenicity in both Ink4a/Arf-dependent and independent manners in mice. Hepatology 2010;52:1111- 23.
- 82. Xu CR, Lee S, Ho C, Bommi P, Huang SA, Cheung ST, Dimri GP, Chen X. Bmi1 functions as an oncogene independent of Ink4A/Arf repression in hepatic carcinogenesis. Mol Cancer Res 2009;7:1937-45.
- 83. Yonemitsu Y, Imazeki F, Chiba T, Fukai K, Nagai Y, Miyagi S, Arai M, Aoki R, Miyazaki M, Nakatani Y, Iwama A, Yokosuka O. Distinct expression of polycomb group proteins EZH2 and BMI1 in hepatocellular carcinoma. Hum Pathol 2009;40:1304-11.
- 84. Chiba T, Miyagi S, Saraya A, Aoki R, Seki A, Morita Y, Yonemitsu Y, Yokosuka O, Taniguchi H, Nakauchi H, Iwama A. The polycomb gene product BMI1 contributes to the maintenance of tumor-initiating side population cells in hepatocellular carcinoma. Cancer Res 2008;68:7742-9.
- 85. Li W, Li Y, Tan Y, Ma K, Cui J. Bmi-1 is critical for the proliferation and invasiveness of gastric carcinoma cells. J Gastroenterol Hepatol 2010;25:568-75.
- 86. Liu JH, Song LB, Zhang X, Guo BH, Feng Y, Li XX, Liao WT, Zeng MS, Huang KH. Bmi-1 expression predicts prognosis for patients with gastric carcinoma. J Surg Oncol 2008;97:267-72.
- 87. Douglas D, Hsu JH, Hung L, Cooper A, Abdueva D, van Doorninck J, Peng G, Shimada H, Triche TJ, Lawlor ER. BMI-1 promotes Ewing sarcoma tumorigenicity independent of CDKN2A repression. Cancer Res 2008;68:6507- 15.
- 88. Li DW, Tang HM, Fan JW, Yan DW, Zhou CZ, Li SX, Wang XL, Peng ZH. Expression level of Bmi-1 oncoprotein is associated with progression and prognosis in colon cancer. J Cancer Res Clin Oncol 2010;136:997-1006.
- 89. Qin ZK, Yang JA, Ye YL, Zhang X, Xu LH, Zhou FJ, Han H, Liu ZW, Song LB, Zeng MS. Expression of Bmi-1 is a prognostic marker in bladder cancer. BMC Cancer 2009;9:61.
- 90. He XT, Cao XF, Ji L, Zhu B, Lv J, Wang DD, Lu PH, Cui HG. Association between Bmi1 and clinicopathological status of esophageal squamous cell carcinoma. World J Gastroenterol 2009;15:2389-94.
- 91. Sasaki M, Yamaguchi J, Ikeda H, Itatsu K, Nakanuma Y. Polycomb group protein Bmi1 is overexpressed and essential in anchorage-independent colony formation, cell proliferation and repression of cellular senescence in cholangiocarcinoma: tissue and culture studies. Hum

Pathol 2009;40:1723-30.

- 92. Honig A, Weidler C, Häusler S, Krockenberger M, Buchholz S, Köster F, Segerer SE, Dietl J, Engel JB. Overexpression of polycomb protein BMI-1 in human specimens of breast, ovarian, endometrial and cervical cancer. Anticancer Res 2010;30:1559-64.
- 93. Michael LE, Westerman BA, Ermilov AN, Wang A, Ferris J, Liu J, Blom M, Ellison DW, van Lohuizen M, Dlugosz AA. Bmi1 is required for Hedgehog pathwaydriven medulloblastoma expansion. Neoplasia 2008;10: 1343-9.
- 94. Zakrzewska M, Zakrzewski K, Grešner SM, Piaskowski S, Zalewska-Szewczyk B, Liberski PP. Polycomb genes expression as a predictor of poor clinical outcome in children with medulloblastoma. Childs Nerv Syst 2011;27:79- 86.
- 95. Leung C, Lingbeek M, Shakhova O, Liu J, Tanger E, Saremaslani P, Van Lohuizen M, Marino S. Bmi1 is essential for cerebellar development and is overexpressed in human medulloblastomas. Nature 2004;428:337-41.
- 96. Hoenerhoff MJ, Chu I, Barkan D, Liu ZY, Datta S, Dimri GP, Green JE. BMI1 cooperates with H-RAS to induce an aggressive breast cancer phenotype with brain metastases. Oncogene 2009;28:3022-32.
- 97. Boukarabila H, Saurin AJ, Batsché E, Mossadegh N, van Lohuizen M, Otte AP, Pradel J, Muchardt C, Sieweke M, Duprez E. The PRC1 Polycomb group complex interacts with PLZF/RARA to mediate leukemic transformation. Genes Dev 2009;23:1195-206.
- 98. Rizo A, Horton SJ, Olthof S, Dontje B, Ausema A, van Os R, van den Boom V, Vellenga E, de Haan G, Schuringa JJ. BMI1 collaborates with BCR-ABL in leukemic transformation of human CD34+ cells. Blood 2010;116:4621- 30.
- 99. Li J, Gong LY, Song LB, Jiang LL, Liu LP, Wu J, Yuan J, Cai JC, He M, Wang L, Zeng M, Cheng SY, Li M. Oncoprotein Bmi-1 renders apoptotic resistance to glioma cells through activation of the IKK-nuclear factor-kappaB Pathway. Am J Pathol 2010;176:699-709.
- 100. Watanabe O, Imamura H, Shimizu T, Kinoshita J, Okabe T, Hirano A, Yoshimatsu K, Konno S, Aiba M, Ogawa K. Expression of twist and wnt in human breast cancer. Anticancer Res 2004;24:3851-6.
- 101. Lee TK, Poon RT, Yuen AP, Ling MT, Kwok WK, Wang XH, Wong YC, Guan XY, Man K, Chau KL, Fan ST. Twist overexpression correlates with hepatocellular carcinoma metastasis through induction of epithelial-mesenchymal transition. Clin Cancer Res 2006;12:5369-76.
- 102. Yan-Qi Z, Xue-Yan G, Shuang H, Yu C, Fu-Lin G, Fei-Hu B, Shi-Ren S, Xu-Feng W, Jie-D, Dai-Ming F. Expression and significance of TWIST basic helix-loophelix protein over-expression in gastric cancer. Pathology 2007;39:470-5.
- 103.Rosivatz E, Becker I, Specht K, Fricke E, Luber B, Busch R, Höfler H, Becker KF. Differential expression of

the epithelial-mesenchymal transition regulators snail, SIP1, and twist in gastric cancer. Am J Pathol 2002;161:1881-91.

- 104. Yuen HF, Chua CW, Chan YP, Wong YC, Wang X, Chan KW. Significance of TWIST and E-cadherin expression in the metastatic progression of prostatic cancer. Histopathology 2007;50:648-58.
- 105. Hung JJ, Yang MH, Hsu HS, Hsu WH, Liu JS, Wu KJ. Prognostic significance of hypoxia-inducible factor-1a, TWIST1, and Snail expression in resectable non-small cell lung cancer. Thorax 2009;64:1082-9.
- 106. Valdés-Mora F, Gómez del Pulgar T, Bandrés E, Cejas P, Ramírez de Molina A, Pérez-Palacios R, Gallego-Ortega D, García-Cabezas MA, Casado E, Larrauri J, Nistal M, González-Barón M, García-Foncillas J, Lacal JC. TWIST1 overexpression is associated with nodal invasion and male sex in primary colorectal cancer. Ann Surg Oncol 2009;16:78-87.
- 107. Wallerand H, Robert G, Pasticier G, Ravaud A, Ballanger P, Reiter RE, Ferrière JM. The epithelial-mesenchymal transition-inducing factor TWIST is an attractive target in advanced and/or metastatic bladder and prostate cancers. Urol Oncol 2010;28:473-9.
- 108. Hasselblatt M, Mertsch S, Koos B, Riesmeier B, Stegemann H, Jeibmann A, Tomm M, Schmitz N, Wrede B, Wolff JE, Zheng W, Paulus W. TWIST-1 is overexpressed in neoplastic choroid plexus epithelial cells and promotes proliferation and invasion. Cancer Res 2009;69:2219-23.
- 109. Waldmann J, Slater EP, Langer P, Buchholz M, Ramaswamy A, Walz MK, Schmid KW, Feldmann G, Bartsch DK, Fendrich V. Expression of the transcription factor snail and its target gene Twist are associated with malignancy in pheochromocytomas. Ann Surg Oncol 2009;16:1997-2005.
- 110. Yoshida J, Horiuchi A, Kikuchi N, Hayashi A, Osada R, Ohira S, Shiozawa T, Konishi I. Changes in the expression of E-cadherin repressors, Snail, Slug, SIP1, and Twist, in the development and progression of ovarian carcinoma: the important role of Snail in ovarian tumorigenesis and progression. Med Mol Morphol 2009;42:82- 91.
- 111. Xie F, Li K, Ouyang X. Twist, an independent prognostic marker for predicting distant metastasis and survival rates of esophageal squamous cell carcinoma patients. Clin Exp Metastasis 2009;26:1025-32.
- 112. Sun T, Zhao N, Zhao XL, Gu Q, Zhang SW, Che N, Wang XH, Du J, Liu YX, Sun BC. Expression and functional significance of Twist1 in hepatocellular carcinoma: its role in vasculogenic mimicry. Hepatology 2010;51: 545-56.
- 113. Okada T, Suehiro Y, Ueno K, Mitomori S, Kaneko S, Nishioka M, Okayama N, Sakai K, Higaki S, Hazama S, Hirata H, Sakaida I, Oka M, Hinoda Y. TWIST1 hyper-

methylation is observed frequently in colorectal tumors and its overexpression is associated with unfavorable outcomes in patients with colorectal cancer. Genes Chromosomes Cancer 2010;49:452-62.

- 114. Li X, Marcondes AM, Gooley TA, Deeg HJ. The helixloop-helix transcription factor TWIST is dysregulated in myelodysplastic syndromes. Blood 2010;116:2304-14.
- 115. Yu Q, Zhang K, Wang X, Liu X, Zhang Z. Expression of transcription factors snail, slug, and twist in human bladder carcinoma. J Exp Clin Cancer Res 2010;29:119.
- 116. Tjensvoll K, Oltedal S, Farmen RK, Shammas FV, Heikkilä R, Kvaløy JT, Gilje B, Smaaland R, Nordgård O. Disseminated tumor cells in bone marrow assessed by TWIST1, cytokeratin 19, and mammaglobin A mRNA predict clinical outcome in operable breast cancer patients. Clin Breast Cancer 2010;10:378-84.
- 117. Peinado H, Ballestar E, Esteller M, Cano A. Snail mediates E-cadherin repression by the recruitment of the Sin3A/histone deacetylase 1 (HDAC1)/HDAC2 complex. Mol Cell Biol 2004;24:306-19.
- 118. Hou Z, Peng H, Ayyanathan K, Yan KP, Langer EM, Longmore GD, Rauscher FJ 3rd. The LIM protein AJUBA recruits protein arginine methyltransferase 5 to mediate SNAIL-dependent transcriptional repression. Mol Cell Biol 2008;28:3198-207.
- 119. Herranz N, Pasini D, Díaz VM, Francí C, Gutierrez A, Dave N, Escrivà M, Hernandez-Muñoz I, Di Croce L, Helin K, Garcia de Herreros A, Peiro S. Polycomb complex 2 is required for E-cadherin repression by the Snail1 transcription factor. Mol Cell Biol 2008;28:4772-81.
- 120. Ansieau S, Bastid J, Doreau A, Morel AP, Bouchet BP, Thomas C, Fauvet F, Puisieux I, Doglioni C, Piccinin S, Maestro R, Voeltzel T, Selmi A, Valsesia-Wittmann S, Caron de Fromentel C, Puisieux A. Induction of EMT by Twist proteins as a collateral effect of tumor-promoting inactivation of premature senescence. Cancer Cell 2008;14:79-89.
- 121. Dimri GP, Martinez JL, Jacobs JJ, Keblusek P, Itahana K, Van Lohuizen M, Campisi J, Wazer DE, Band V. The Bmi-1 oncogene induces telomerase activity and immortalizes human mammary epithelial cells. Cancer Res 2002;62:4736-45.
- 122. Kim RH, Lieberman MB, Lee R, Shin KH, Mehrazarin S, Oh JE, Park NH, Kan MK. Bmi-1 extends the life span of normal human oral keratinocytes by inhibiting the TGF-beta signaling. Exp Cell Res 2010;316:2600-8.
- 123. Song LB, Li J, Liao WT, Feng Y, Yu CP, Hu LJ, Kong QL, Xu LH, Zhang X, Liu WL, Li MZ, Zhang L, Kang TB, Fu LW, Huang WL, Xia YF, Tsao SW, Li M, Band V, Band H, Shi QH, Zeng YX, Zeng MS. The polycomb group protein Bmi-1 represses the tumor suppressor PTEN and induces epithelial-mesenchymal transition in human nasopharyngeal epithelial cells. J Clin Invest 2009;119:3626-36.

Twist1 直接活化 Bmi1 基因的表達:對癌症幹細胞特性, 上皮細胞一間質細胞轉化,及臨床癌症的重要性

吳國瑞

癌症幹細胞的觀念可用來描述一小群存在腫瘤中具有幹細胞特性的癌細胞。癌症幹細胞 具有自我更新的能力並且可以抵抗化療和輻射治療。上皮細胞—間質細胞轉化是癌症轉移的 重要機制,此過程讓癌細胞具有幹細胞持性。然而上皮細胞––間質細胞轉化和癌症幹細胞的 產生兩者間的關係目前並沒有詳細的機制可以解釋。Bmi1 是 polycomb repressive complex 1 (PRC1) 之一員。Bmi1 的功能在於可以維持自我更新能力和幹細胞特性。目前研究指出上皮細 胞–間質細胞轉化調控分子 Twist1 可以直接活化 Bmi1 基因的表達,這兩個分子須互相依存 進而共同調控上皮細胞一間質細胞轉化和癌症幹細胞特性。這個結果可以解釋上皮細胞一間 質細胞轉化和癌症幹細胞特性兩者的關係。Bmi1 大量表現於人類的各種癌症,並且讓癌細胞 具有抗藥性。Twist1 也大量表現在人類的各種癌症並且可做為預測病人預後的標記。Twist1 和 Bmi1 功能性的互相依存與交互作用提供解釋上皮細胞––間質細胞轉化進而誘導癌症幹細 胞特性一個嶄新的概念。未來進一步探討上皮細胞––間質細胞轉化誘導癌症幹細胞特性的機 制可以對轉移性癌症的處理及治療有更顯著的貢獻。(長庚醫誌 2011;34:229-38)

關鍵詞: 上皮細胞─間質轉化,癌症幹細胞,癌症轉移, Twist1, Bmi1